Biomechanical Properties and Mechanotransduction of Skeletal Muscles as The Basis for The Use of Myofascial Osteopathic Techniques

S. V. Novoseltsev^{1*}, A. G. Reshetnikov², V. D. Vyunov³, T. R. Suleymanov⁴, A. M. Nefedova⁵, P. G. Solomatov⁶, S. I. Taranenko⁷, A. M. Kryazheva⁸, S. V. Moskvicheva⁹

¹Department of Sports Medicine and Medical Rehabilitation of the Institute of Clinical Medicine. N.V. Sklifosovsky Federal State Autonomous Educational Institution of Higher Education First Moscow State Medical University. THEM. Sechenov of the Ministry of Health of Russia; E-mail: <u>snovoselcev@mail.ru</u>

^{2,3,4,5,6,7}Doctor osteopath, Department of Sports Medicine and Medical Rehabilitation, I.M. Sechenov First Moscow State Medical University (Sechenov University) (Moscow, Russia)

⁸Resident physician in osteopathy, Department of Sports Medicine and Medical Rehabilitation, I.M. Sechenov First Moscow State Medical University (Sechenov University) (Moscow, Russia)

⁹Doctor osteopath, «M+Clinic» Medical Center 142440, 15 Yakovleva Str., Obuhovo, Noginsk district, Moscow rgn., Russian Federation

Abstracts: Skeletal muscles are an important link in human biomechanics. Every movement depends on muscle strength and is important for maintaining health and quality of life. Mechanical loads on the human body directly depend on the state of muscle tissue, so it is necessary to understand the cellular composition and regulation of the tissue responsible for the processes of mechanotransduction. This, in turn, makes it possible to develop methods of effective manual intervention in complex rehabilitation for patients with dysfunctions of the musculoskeletal system. This review describes the molecular basis of mechanotransduction in muscle tissue.

Keywords: Hypertrophy, mTORC, Wnt Growth, Mechanical Load, Mechanotransduction, Skeletal Muscles.

1. INTRODUCTION

Any movement is carried out by our skeletal muscles. We are used to thinking that a muscle can contract and stretch, but a large number of chemical reactions take place in these tissues. This is an important component in human physiology, providing a greater range of motion in joints and improving microcirculation of all organs. For example, a number of chemical reactions occur in skeletal muscle, as well as the storage of amino acids and glucose. Produces various myokines that influence metabolism in other tissues [1-3].

We must understand the factors that influence skeletal muscle metabolism to prevent loss of muscle mass, which can lead to functional instability and increase the risk of injury and ultimately reduce quality of life.

A huge number of factors influence muscles, both genetic and epigenetic: hormonal levels, nutrition, mechanical stimuli [4,5]. Mechanical loading is necessary for the growth of muscle tissue (i.e., muscle hypertrophy) [6-8].

The structural unit of skeletal muscles is muscle fiber and the application of a mechanical load on it will give a biochemical reaction, the so-called process is mechanotransduction

Muscle fiber consists of myofibrils, which include molecules <u>actin</u> and <u>myosin</u>, as well as auxiliary proteins - <u>titin</u>, <u>troponin</u>, <u>tropomyosin</u> and other proteins. All these components are found in sarcaplasm.

When skeletal muscles contract, sarcoplasm, like water in a balloon, presses on the outer shell <u>muscle fiber</u> and expands it, activating chemical signals inside <u>fibers</u>. Chemical signals activate a variety of enzymes found in the

sarcoplasm. These enzymes act on the myonuclei and cause increased protein synthesis. This ultimately leads to <u>muscle hypertrophy.</u>

The lack of muscle function even in the womb leads to impaired development of fetal bones and joints [13-17], and during postnatal skeletal growth (2-20 years), the more muscle mass increases, the more bone mass becomes [18]. If we compare muscle and bone tissue, we can say with confidence that they are similar [19, 20]. This is how muscles work to grow bones. And a decrease in muscle mass can lead to bone loss, which will lead to spinal cord damage [23], and the development of neuromuscular diseases (cerebral palsy, Duchenne muscular dystrophy [21, 22].

In the practice of a manual medicine doctor, dysfunctions of the musculoskeletal system are often encountered, caused by weakness of skeletal muscles, with the formation of local muscle compactions (triggers) in them. To eliminate them, myofascial release techniques are used. release, soft tissue, muscle-energy techniques, strain - counterstrain techniques. In addition to manual treatment, special exercises and a corrective diet are prescribed to consolidate the therapeutic effect. However, the doctor does not always fully understand the specifics of the physiological processes occurring in muscle tissue as a result of manual mechanical action. In this article, we focused on mechanotransduction, a physiological process in which mechanical load is converted into biochemical reactions.

1.1. Mechanotransduction And Regulation of Skeletal Muscle Mass

The development of muscle tissue depends on protein synthesis and degradation [24]. The greater the synthesis, the stronger the muscle hypertrophy, and conversely, reduced synthesis leads to muscle atrophy. Currently, there are a number of signaling mechanisms that influence protein synthesis under mechanical load [25-34]. For example, a protein kinase called mechanical target of rapamycin (mTOR), [6, 11]. Also, satellite cells (SC) play an important role in mechanotransduction [7, 35 - 38].

1.2. Mechanical Signaling Messengers in Skeletal Muscle

mTOR

mTOR is a conserved serine / threonine kinase that is contained in two complexes: mTORC1 (whose signaling is partially inhibited by the drug rapamycin) and mTORC2 (resistant to rapamycin) [39]. mTORC1 is the main regulator of cell growth, ensures the rate of mRNA translation), and regulates the number of ribosomes [40].

Under the influence of mechanical forces, mTORC1 is activated in skeletal muscles [6]. As a result, protein synthesis increases and induces hypertrophy of muscle fibers (*in vivo*) [41, 42], which lead to muscle hypertrophy [43, 44].

Potential candidates for mechanically induced mTORC1 activation include increased intracellular Ca2, amino acids, insulin-like growth factor 1 (IGF -1), extracellular signal-regulated kinase 1 and 2 (ERK1/2), phosphatidic acid (PA), TSC2 (tuberin). Additional research is needed to clarify the role of the above factors.

1.3. Increased Intracellular Calcium Content

The sarcoplasmic reticulum (SR) stimulates the release of calcium, which causes skeletal muscle contraction [45]. There is also a downside - with passive muscle stretching, the amount of intracellular calcium increases by activating stretch-sensitive ion channels [46, 47]. Studies in non-muscle cells have shown that mTORC1 signaling can be regulated by changes in [Ca2⁺], with an increase in [Ca2⁺] activating mTORC1 signaling and a decrease in [Ca2⁺] resulting in inhibition of mTORC1 signaling [48, 49]. Increasing [Ca2⁺] has been shown to also increase the rate of protein synthesis *ex vivo* in resting skeletal muscle [50]. Thus, contraction or stretch induced increases in [Ca2⁺] appear to be an ideal candidate for mechanical activation of mTORC1 and protein synthesis in muscle tissue.

Chronic mechanical overload activates Ca2 ⁺ / calmodulin-dependent protein kinase α (CaMKK α), and together with the increase in Ca2 ⁺ stimulates mTORC1 signaling and protein synthesis [52, 53]. On the other hand, there is no requirement for CaMKK α for muscle growth [53].

Thus, an increase in [Ca2+] may stimulate mechanical activation of mTORC1, but this requires further study.

1.4. Amino Acids

Amino acids are important regulators of mTORC1 signaling, which plays a key role in the regulation of protein synthesis and skeletal muscle mass. Studies have shown that decreasing amino acid levels leads to decreased mTORC1 activation, while increasing amino acid availability increases mTORC1 activation [60].

Particularly important are branched chain amino acids such as leucine, which can activate mTORC1 signaling and increase protein synthesis. Therefore, increased amino acid intake may play a role in the mechanical activation of mTORC1 signaling [61, 62].

However, there is still no evidence to support this hypothesis. Some studies have shown that resistance exercise increases muscle leucine content and class III PI3K, Vps34 activity, which is associated with mTORC1 activation [63 - 67]. However, other studies have reported a decrease in leucine following acute resistance exercise [68].

Thus, although there is some indirect evidence suggesting a role for amino acids in the mechanical activation of mTORC1 signaling, more research is required to more fully understand this process.

1.5. Prostaglandins

There is evidence that prostaglandin (PG) signaling may also play a role in the mechanical regulation of protein synthesis and muscle hypertrophy. Studies have shown that PGs stimulate an increase in protein synthesis in skeletal muscle and cause hypertrophy of cultured myotubes [69 - 74].

In addition, mechanical stimulation of skeletal muscle also leads to increased production of PGs, including PGF2α [75, 76]. This supports a possible role for PG in mechanical activation of mTORC1 and muscle hypertrophy. It is also worth mentioning that after eccentric exercise there is an increase in protein synthesis, and large doses of Cox inhibitors, acetaminophen and ibuprofen can reduce it [77].

Not all PGs can stimulate protein synthesis. Some studies have shown that PGE2 stimulates protein degradation in skeletal muscle [70]. Therefore, additional research is required to more fully understand the role of various PGs in the mechanical regulation of protein synthesis and skeletal muscle mass.

Overall, although there is some evidence suggesting a role for amino acids and prostaglandins in the mechanical regulation of mTORC1 signaling and protein synthesis, more research is required to more fully understand these processes.

1.6. Wnt Signaling

In skeletal muscle embryogenesis as regeneration, Wnt molecules play a major role [78, 79]. It is also worth noting that exercise increases Dsh/GSK3 β interaction and also decreases GSK3 β activity and β - catenin phosphorylation in humans [80]. In addition, muscle fiber hypertrophy induced by resistance training is associated with increased expression of various elements of the Wnt signaling pathway (e.g., Wnt1, β - catenin , LEF1, cyclin D1) [81]. Muscle loading also influences muscle hypertrophy, and β -catenin is required for this [82]. It follows that Wnt/ β - catenin signaling takes place in mechanotransduction and muscle tissue enlargement.

CONCLUSION

In conclusion, we note that the molecular mechanisms regulating mechanotransduction in skeletal muscle are complex and, in many ways, similar to the processes in bone tissue. In this case, several mechanisms likely work in synergy.

In skeletal muscle, mTORC1 has been established as a major messenger for stimulating protein synthesis, as well as intracellular calcium and amino acids. However, much remains to be studied to prove the acceleration of skeletal muscle growth due to mechanotransduction messengers.

The role of prostaglandins and Wnt signaling in the mechanical activation of mTORC1 in skeletal muscle needs to be understood.

This review explains the basic processes of mechanotransduction in skeletal muscle. It is necessary to understand the importance of studying this topic to explain the application of myofascial osteopathic techniques, physical exercises for the development and growth of muscle tissue and stimulate ideas for future research.

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Conflict of interest

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REFERENCES

- [1] Pedersen BK, February, Massachusetts. Muscle, exercise and obesity: skeletal muscle as a secretory organ. Nat Rev Endocrinol. 2012;8:457–465. [PubMed] [Google Scholar]
- [2] Yang J. Enhanced skeletal muscle for efficient glucose homeostasis. In: Ya-Xiong T, editor. Progress in Molecular Biology and Translational Science: Academic Press. 2014. pp. 133-163. [PubMed] [Google Scholar]
- [3] Wolfe R.R. The role of muscles in health and disease is underestimated. I Jay Wedge Nutr . 2006;84:475–482. [PubMed] [Google Scholar]
- [4] Schiaffino S, Dyar KA, Ciciliot S, Blaauw B, Sandri M. Mechanisms regulating height skeletal muscles And atrophy. FEBS Magazine . 2013; 280:4294–4314. [PubMed] [Google Scholar]
- [5] Robling AG. Interaction of biological factors with mechanical signals in bone adaptation: recent developments. Curr Osteoporos Rep . 2012;10:126-131. [PMC free article] [PubMed] [Google Scholar]
- [6] Goodman, California. The role of mTORC1 in regulating protein synthesis and skeletal muscle mass in response to various mechanical stimuli. Rev Physiol Biochem Pharmacol . 2014; 166:1–53. [PubMed] [Google Scholar]
- [7] Adams G.R., Bamman M.M. Characterization and regulation of mechanical compensatory muscle hypertrophy caused by mechanical load. Comprehensive Physiology. 2012;2:2829–2870. [PubMed] [Google Scholar]
- [8] Hornberger T.A. Mechanotransduction and regulation of mTORC1 signaling in skeletal muscle. Int J Biochem Cell Biol. 2011;43:1267–1276. [Free PMC article] [PubMed] [Google Scholar]
- [9] Arfat Y, Xiao WZ, Iffikhar S, Zhao F, Li DJ, Sun YL, Zhang G, Shang P, Qian AR. Physiological effects of microgravity on bone cells. International Calcified Tissue Organization. 2014;94:569-579. [PubMed] [Google Scholar]
- [10] Klein- Nulend J, Bacabac RG, Bakker AD. Mechanical load and how it affects bone cells: the role of the osteocyte cytoskeleton in maintaining our skeleton. Eur Cell Mater . 2012;24:278–291. [PubMed] [Google Scholar]
- [11] Robling AG, Turner CH. Mechanical signaling for bone modeling and reconstruction. Cirt Rev Eukaryot Gene Expr. 2009;19:319-338. [Free PMC article] [PubMed] [Google Scholar]
- [12] Lu TW, Taylor SJ, O'Connor JJ, Walker PS. The influence of muscle activity on femoral forces: a study in vivo. J. Biomech . 1997;30:1101– 1106. [PubMed] [Google Scholar]
- [13] Rodriguez G, Garcia-Alix A, Palacios J, Paniagua R. Changes in long bones due to fetal immobility caused by neuromuscular diseases. Xray and histological examination. J Bone Joint Surg Am. 1988;70:1052–1060. [PubMed] [Google Scholar]

- [14] Rodriguez H, Palacios H, Garcia-Alix A, Pastor I, Paniagua R. Effect of immobilization on fetal bone development. Morphometric study in newborns with congenital neuromuscular diseases with intrauterine disease. International Calcified Tissue Organization. 1988;43:335–339. [PubMed] [Google Scholar]
- [15] Sharir A, Stern T, Gnil S, Shahar R, Seltzer E. Muscle force regulates bone formation for optimal load-bearing capacity during embryogenesis. Development. 2011;138:3247–3259. [PubMed] [Google Scholar]
- [16] Hall BC, Herring SW. Paralysis and growth of the musculoskeletal system in the embryonic chick. Diary of morphology. 1990; 206:45–56. [PubMed] [Google Scholar]
- [17] Hall JG. The importance of muscle movement for normal craniofacial life. J Craniofac Surg . 2010;21:1336-1338. [PubMed] [Google Scholar]
- [18] Rauch F, Bailey DA, Baxter-Jones A, Mehrwald R, Faulkner R. "Muscle-bone" during the pubertal growth spurt. Bone. 2004;34:771-775. [PubMed] [Google Scholar]
- [19] Rauch F, Schonau E. Peripheral quantitative computed tomography of the proximal radius in young subjects new reference data and interpretation of results. J Musculoskelet Neuronal Interact. 2008;8:217–226. [PubMed] [Google Scholar]
- [20] Fricke O, Becquart R, Semler O, Schonau E. Analysis of muscle mass and function: effects on bone mineral density and peak muscle mass. Pediatrician Nephrol . 2010; 25:2393–2400. [PubMed] [Google Scholar]
- [21] Larson SM, Henderson RC. Bone density and fractures in boys with Duchenne muscular dystrophy. J. Pediatrician Orthop. 2000;20:71–74. [PubMed] [Google Scholar]
- [22] Tasdemir HA, Buyukavci M, Akcay F, Polat P, Yildiran A, Karakelleoglu C. Bone mineral density in children with cerebral palsy. International Pediatrics. 2001;43:157–160. [PubMed] [Google Scholar]
- [23] Eser P, Frotzler A, Zehnder Y, Wick L, Knecht H, Denoth J, Schiessl H. Association between duration of paralysis and bone structure: a pQCT study of people with spinal cord injury. Bone. 2004;34:869-880. [PubMed] [Google Scholar]
- [24] Goodman, California, Mayhew D.L., Hornberger, T.A. Recent progress in understanding the molecular mechanisms regulating skeletal muscle mass. Cellular signal. 2011;23:1896–1906. [PMC free article] [PubMed] [Google Scholar]
- [25] Lynch G.S., Ryall J.G. The role of {beta}-adrenergic receptor signaling in skeletal muscle: implications for muscle wasting and disease. Physiol. Ed. 2008; 88:729-767. [PubMed] [Google Scholar]
- [26] Glass DJ. PI3 kinase regulation of skeletal muscle hypertrophy and atrophy. Curr Top Microbiol Immunol. 2010;346 [PubMed] [Google Scholar]
- [27] Huang Z, Chen X, Chen D. Myostatin: new insights into its role in metabolism, signaling pathways and expression regulation. Cellular signal. 2011;23:1441–1446. [PubMed] [Google Scholar]
- [28] Lee N.K., McLean. Polyamines, androgens and skeletal muscle hypertrophy. J Cell Physiol . 2011;226:1453–1460. [PubMed] [Google Scholar]
- [29] McCarthy JJ. MyomiR network in skeletal muscle plasticity. Exerc Sport Sci Rev. 2011;39:150-154. [Free PMC article] [PubMed] [Google Scholar]
- [30] Phillips BE, Hill DS, Atherton PJ. Regulation of muscle protein synthesis in humans. Curr Opin Clin Nutr Metab Care. 2012;15:58-63. [PubMed] [Google Scholar]
- [31] Schiafino S, Mammukari S. Regulation of skeletal muscle growth by IGF1-Akt/PKB: insights from genetic models. Skeletal muscle. 2011;1:4. [PMC free article] [PubMed] [Google Scholar]
- [32] Berdo R, Stewart R. cAMP signaling in skeletal muscle adaptation: hypertrophy, metabolism and regeneration. American Journal of Physiology - Endocrinology and Metabolism. 2012;303:E1-E17. [PMC free article] [PubMed] [Google Scholar]
- [33] Dubois V, Laurent M, Boonen S, Vanderschuren D, Klassen F. Androgens and skeletal muscle: cellular and molecular mechanisms underlying anabolic actions. Cellular and molecular life sciences. 2012;69:1651–1667. [PubMed] [Google Scholar]
- [34] Piccirillo R, Demontis F, Perrimon N, Goldberg A.L. Mechanisms of muscle growth and atrophy in mammals and Drosophila. Dynamics of development. 2013 in the press. [PMC free article] [PubMed] [Google Scholar]
- [35] Blau B, Regiani S. The role of satellite cells in muscle hypertrophy. J Muscle Res Cell Motil . 2014;35:3–10. [PubMed] [Google Scholar]
- [36] McCarthy JJ, Mula J, Miyazaki M, Erfani R, Garrison K, Farooqui AB, Srikuea R, Lawson BA, Grimes B, Keller C, Van Zant G, Campbell KS, Esser KA, Dupont- Versteegden EE, Peterson CA. Effective fiber hypertrophy in satellite cell-depleted skeletal muscle. Development. 2011;138:3657–3666. [Free PMC article] [PubMed] [Google Scholar]
- [37] Fornaro M, Hinken AC, Needle S, Hu E, Trendelenburg AU, Mayer A, Rosenstiel A, Chang C, Meier V, Billin AN, Becherer JD, Brace AD, Evans WJ, Glass DJ, Russell AJ. Mechanogrowth factor (MGF) peptide has no apparent effect on muscle myoblasts or primary muscle stem cells. Am J Physiol Endocrinol Metab . 2013 in press. [PubMed] [Google Scholar]
- [38] O'Connor RS, Pavlath GK, McCarthy JJ, Esser KA. Last word on point: Counterpoint: Adding a satellite cell is/isn't necessary for skeletal muscle hypertrophy. J Appl Physiol . 2007;103:1107. [PubMed] [Google Scholar]
- [39] Laplante M, Sabatini David M. mTOR signaling in growth control and disease. Cell. 2012;149:274–293. [PMC free article] [PubMed] [Google Scholar]
- [40] Mahoney S.J., Dempsey J.M., Blenis J. Cell signaling in protein synthesis, ribosome biogenesis and translation initiation and elongation. Prog Mol Biol Transl Sci. 2009; 90C: 53-107. [PubMed] [Google Scholar]
- [41] Goodman CA, Mabrey DM, Frey JW, Miu MH, Schmidt EK, Pierre P, Hornberger TA. New insights into the regulation of skeletal muscle

protein synthesis revealed by a new non-radioactive technique in vivo . FACEB J. 2011;25:1028–1039. [PMC free article] [PubMed] [Google Scholar]

- [42] Goodman CA, Miu MH, Frey JW, Mabrey DM, Lincoln HC, Ge Y, Chen J, Hornberger TA. Phosphatidylinositol 3-kinase/ protein kinase Bindependent activation of the mammalian target of rapamycin is sufficient to induce skeletal muscle hypertrophy. Mol. Biol. Cell. 2010;21:3258–3268. [PMC free article] [PubMed] [Google Scholar]
- [43] Bodine SC, Stitt TN, Gonzalez M, Kline WO, Stover G.L. Bauerlein R, Zlotchenko E, Scrimgeour A, Lawrence J.C. Glass DJ, Yancopoulos G.D. The Akt / mTOR pathway is a critical regulator of skeletal muscle hypertrophy and may prevent muscle atrophy in vivo. Nat Cell Biol. 2001;3:1014–1019. [PubMed] [Google Scholar]
- [44] Goodman CA, Frey JW, Mabrey DM, Jacobs BL, Lincoln HC, You JS, Hornberger TA. The role of skeletal muscle mTOR in the regulation of mechanical load-induced growth. J Physiol. 2011;589:5485–5501. [Free PMC article] [PubMed] [Google Scholar]
- [45] Stephenson DG, Lamb GD, Stephenson GM. Excitation-contraction-relaxation (E-CR) cycle events in mammalian muscle fibers relevant to muscle fatigue. Acta Physiol Scand . 1998;162:229–245. [PubMed] [Google Scholar]
- [46] Armstrong RB, Duan C, Delp MD, Hayes DA, Glenn GM, Allen GD. Increases in plantar muscles of rats [Ca2+] with passive stretch. J Appl Physiol . 1993;74:2990–2997. [PubMed] [Google Scholar]
- [47] Franco A., Lansman J.B. Stretch-sensitive channels in muscle cell development from a mouse cell line. Journal of Physiology. 1990; 427:361–380. [PMC free article] [PubMed] [Google Scholar]
- [48] Graves LM, He Y, Lambert JHunter D, Li X, Earp H.S. Intracellular calcium signal activates p70 but not p90 ribosomal S6 kinase in liver epithelial cells. Magazine biological chemistry 1997;272:1920–1928. [PubMed] [Google Scholar]
- [49] Conus NM, Hemmings BA, Pearson RB. Differential calcium regulation reveals distinct signaling requirements for Akt and p70S6k activation. J Biol. Chemistry. 1998;273:4776–4782. [PubMed] [Google Scholar]
- [50] Kameyama T., Etlinger J.D. Calcium regulation of protein synthesis and degradation in muscles. Nature. 1979;279:344–346. [PubMed] [Google Scholar]
- [51] Ito N, Ruegg UT, Kudo A, Miyagoe -Suzuki Y, Takeda Si. Activation of calcium signaling through Trpv1 by nNOS and peroxynitrite as a key trigger of skeletal muscle hypertrophy. Nat Med. 2013;19:101–106. [PubMed] [Google Scholar]
- [52] McGee SL, Mustard KJ, Hardie DG, Baar K. Normal hypertrophy accompanied by phosphorescence and activation of AMP-activated protein kinase α1 after overload in LKB1 mice. J. Physiol . 2008;586:1731–1741. [Free PMC article] [PubMed] [Google Scholar]
- [53] Ferey JLA, Brault JJ, Smith CAS, Witczak CA. Constitutive activation of CaMKKα signaling is sufficient but not necessary for mTORC1 activation and mouse skeletal muscle growth. Am J Physiol Endocrinol and Metab. 2014 in press: [Free PMC article] [PubMed] [Google Scholar]
- [54] Corradetti, Minnesota, Guan, CL. Upstream of mammalian target of rapamycin: do all roads pass through mTOR? Oncogene. 2006;25:6347–6360. [PubMed] [Google Scholar]
- [55] Powers SC, Jackson MJ. Exercise-induced oxidative stress: cellular mechanisms and effects on muscle force production. Physiol. Rev. _ 2008;88:1243-1276. [PMC free article] [PubMed] [Google Scholar]
- [56] Smith L. W. Smith JD, Criswell D.S. _ Involvement of nitric oxide synthase in the adaptation of skeletal muscles to chronic overload. J Appl Physiol . 2002;92:2005–2011. [PubMed] [Google Scholar]
- [57] Soltow QA, Betters JL, Sellman JE, Lira VA, Long JH, Criswell DS. Ibuprofen inhibits skeletal muscle hypertrophy in rats. Med-scientific sports exercises . 2006;38:840-846. [PubMed] [Google Scholar]
- [58] Soltow QA, Zeanah EH, Lira VA, Criswell DS. The stress of cyclic stretching causes atrophy of C2C12 myotubes. Biochem Biophys Res Commun. 2013;434:316-321. [PubMed] [Google Scholar]
- [59] Suzuki N, Motohashi N, Uezumi A, Fukada S- i, Yoshimura T, Itoyama Y, Aoki M, Miyagoe -Suzuki Y, Takeda Si. NO production leads to suspension-induced muscle atrophy by dislocation of NOS neurons. J. Klin. Invest. 2007;117:2468–2476. [PMC free article] [PubMed] [Google Scholar]
- [60] Jewell J.L., Russell, Guan K-L. The amino acid signals upstream to mTOR. Nat Rev Mol Cell Biol. 2013;14:133–139. [PMC free article] [PubMed] [Google Scholar]
- [61] Anthony JC, Yoshizawa F, Anthony TG, Vari TK, Jefferson LS, Kimball SR. Leucine stimulates translation initiation in skeletal muscle of postabsorptive rats via a rapamycin -sensitive pathway. J. Nutr . 2000;130:2413-2419. [PubMed] [Google Scholar]
- [62] Dickinson JM, Fry CS, Drummond MJ, Gundermann DM, Walker DK, Glynn EL, Timmerman KL, Dhanani S, Volpi E, Rasmussen BB. Activation of mammalian complex 1 to stimulate human skeletal muscle protein synthesis with essential amino acids. J. Nutr . 2011;141:856–862. [PMC free article] [PubMed] [Google Scholar]
- [63] MacKenzie MG, Hamilton DL, Murray JT, Baar K. mVps34 is activated by acute bout of resistance. Biochem Soc Trans. 2007; 035:1314-1316. [PubMed] [Google Scholar]
- [64] Gulati P, Gaspers LD, Dann SG, Joaquin M, Nobukuni T, Natt F, Kozma SC, Thomas AP, Thomas G. Amino acids activate mTOR complex 1 through Ca2+/ CaM signaling on hVps34. Cell metab . 2008;7:456–465. [PMC free article] [PubMed] [Google Scholar]
- [65] Gran P, Cameron-Smith D. Effects of exogenous leucine on signaling and amino acid transporters in human myotubes . Physiology of BMC. 2011;11:10. [PMC free article] [PubMed] [Google Scholar]
- [66] Xu L, Salloum D, Medlin PS, Saqcena M, Yellen P, Perrella B, Foster DA. Phospholipase D mediates nutrient entry into the mammalian target of rapamycin complex 1 (mTORC1) J Biol Chem. 2011;286:25477-25486. [PMC free article] [PubMed] [Google Scholar]

[67] Yoon MS, Du G, Backer JM, Frohman MA, Chen J. Class III PI-3-kinase activates phospholipase D in the mTORC1 amino acid pathway. J-1452 cell biology. 2011;195:435-447. [PMC free article] [PubMed] [Google Scholar]

- [68] D'Souza RF, Marworth JF, Figueiredo VC, Della Gatta PA, Petersen AC, Mitchell CJ, Cameron -Smith D. Dose-dependent increases in p70S6K phosphorylation and intramuscular branched-chain amino acids in older men following resistance exercise and protein intake. Physiol Rep . 2014;2 [PMC free article] [PubMed] [Google Scholar]
- [69] Karim S.M., Sandler M., Williams Ed. Distribution of prostaglandins in human tissues. Br J Pharmacol Chemother. 1967;31:340-344. [Free PMC article] [PubMed] [Google Scholar]
- [70] Rodemann HP, Goldberg AL. Arachidonic acid, prostaglandin E2 and alpha F2 influence the rate of protein turnover in skeletal and cardiac muscle. Journal of Biological Chemistry. 1982;257:1632-1638. [PubMed] [Google Scholar]
- [71] Markworth J.F., Cameron-Smith D. Prostaglandin F2α stimulates PI3K/ERK/ mTOR signaling and skeletal myotube hypertrophy. American Journal of Physiology - Cellular Physiology. 2011; 300: C 671- C 682. [PubMed] [Google Scholar]
- [72] Smith RH, Palmer RM, Reeds PJ. Protein synthesis in isolated rabbit forelimb muscles. Possible role of arachidonic acid metabolites in the intermittent stretch response. Biochemistry. J 1983;214:153-161. [PMC free article] [PubMed] [Google Scholar]
- [73] Markworth J.F., Cameron-Smith D. Arachidonic acid supplementation enhances skeletal cell growth in vitro via a COX-2-dependent pathway. American Journal of Physiology - Cellular Physiology. 2013;304:C56-C67. [PubMed] [Google Scholar]
- [74] Korotkova M., Lundberg I.E. The skeletal muscle arachidonic acid cascade in health and inflammatory diseases. Nat r Reymatol . 2014;10:295-303. [PubMed] [Google Scholar]
- [75] Trappe T.A. Liu S.Z. Effect of prostaglandins and COX inhibitory drugs on the adaptation of skeletal muscles to exercise. Journal of Applied Physiology. 2013; 115:909-919. [PMC free article] [PubMed] [Google Scholar]
- [76] Palmer RM. Prostaglandins and the control of muscle protein synthesis and degradation. Prostaglandins leucote with fatty acids. 1990;39:95–104. [PubMed] [Google Scholar]
- [77] Trappe T.A., White F., Lambert C.P. Cesar D, Hellerstein M Evans W.J. The effect of ibuprofen and acetaminophen on post- exercise muscle protein synthesis. Am J Physiol Endocrinol Metab. 2002; 282:E551-E556. [PubMed] [Google Scholar]
- [78] von Malzahn J, Chang, North Carolina, Benzinger, Rudnitsky Massachusetts. Wnt signals myogenesis. Trends in cell biology. 2012;22:602– 609. [PMC free article] [PubMed] [Google Scholar]

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